

# CS681: Advanced Topics in Computational Biology

Week 8 Lecture 1

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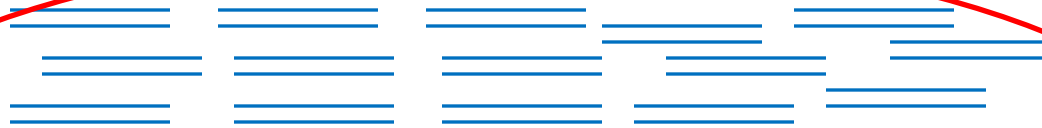
<http://www.cs.bilkent.edu.tr/~calkan/teaching/cs681/>

# Genome Assembly

Test genome



Random shearing and  
Size-selection



Sequencing



Assemble

Contigs/  
scaffolds



# De Bruijn Graphs

- $n$ -dimensional directed graph of  $m$  symbols
  - $m^n$  vertices: all possible length- $n$  sequences of  $m$  symbols
  - Edges between vertices  $v$  and  $w$  if  $sequence(w)$  can be generated by *shifting sequence(v)* by one character and add one new character
  - $S = \{s_1, s_2, \dots, s_m\}$
  - $V = S^n = \{(s_1, \dots, s_1, s_1), (s_1, \dots, s_1, s_2), \dots, (s_m, \dots, s_m, s_m)\}$
  - $E = \{((v_1, v_2, \dots, v_n), (w_1, w_2, \dots, w_n)): v_2=w_1, v_3=w_2, \dots, v_n=w_{n-1}\}$

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# De Bruijn Graph for DNA Assembly

- $m = 4$  (A, C, G, T)
  - $n = k$  (k-mer size)
  - $4^k$  *potential* vertices
    - In reality if  $k$  is sufficiently large, upper bound is genome size
    - Twin vertices: vertices with sequences that are reverse-complement of each other
      - AAAA twin of TTTT
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# De Bruijn Assemblers

- Currently the most common for NGS: Euler, ALLPATHS-LG, Velvet, ABySS, SOAPdenovo
  - Divide reads into k-mers
    - Build graph from k-mers
      - Put an edge if there is k-1 bp prefix-suffix match
    - Error correction
    - Eulerian path
  - The first parts (graph construction & correction) is essentially common to all these assemblers, with a few implementation differences (e.g. parallelization in ABySS)
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# A quick example

**TAGTCGAGGCTTTAGATCCGATGAGGCTTTAGAGACAG**

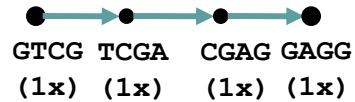
AGTCGAG CTTTAGA CGATGAG CTTTAGA  
GTCGAGG TTAGATC ATGAGGC GAGACAG  
GAGGCTC ATCCGAT AGGCTTT GAGACAG  
AGTCGAG TAGATCC ATGAGGC TAGAGAA  
TAGTCGA CTTTAGA CCGATGA TTAGAGA  
CGAGGCT AGATCCG TGAGGCT AGAGACA  
TAGTCGA GCTTTAG TCCGATG GCTCTAG  
TCGACGC GATCCGA GAGGCTT AGAGACA  
TAGTCGA TTAGATC GATGAGG TTTAGAG  
GTCGAGG TCTAGAT ATGAGGC TAGAGAC  
AGGCTTT ATCCGAT AGGCTTT GAGACAG  
AGTCGAG TTAGATT ATGAGGC AGAGACA  
GGCTTTA TCCGATG TTTAGAG  
CGAGGCT TAGATCC TGAGGCT GAGACAG  
AGTCGAG TTTAGATC ATGAGGC TTAGAGA  
GAGGCTT GATCCGA GAGGCTT GAGACAG

# A quick example

```
AGTCGAG CTTTAGA  CGATGAG CTTTAGA
GTCGAGG  TTAGATC  ATGAGGC   GAGACAG
      GAGGCTC   ATCCGAT AGGCTTT  GAGACAG
AGTCGAG   TAGATCC ATGAGGC  TAGAGAA
TAGTCGA  CTTTAGA  CCGATGA   TTAGAGA
      CGAGGCT  AGATCCG TGAGGCT  AGAGACA
TAGTCGA  GCTTTAG TCCGATG   GCTCTAG
      TCGACGC   GATCCGA  GAGGCTT  AGAGACA
TAGTCGA   TTAGATC GATGAGG  TTTAGAG
      GTCGAGG TCTAGAT  ATGAGGC  TAGAGAC
      AGGCTTT  ATCCGAT AGGCTTT  GAGACAG
AGTCGAG   TTAGATT  ATGAGGC  AGAGACA
      GGCTTTA  TCCGATG   TTTAGAG
      CGAGGCT TAGATCC  TGAGGCT  GAGACAG
AGTCGAG  TTTAGATC  ATGAGGC  TTAGAGA
      GAGGCTT  GATCCGA  GAGGCTT  GAGACAG
```

# A quick example

**First read: GTCGAGG**

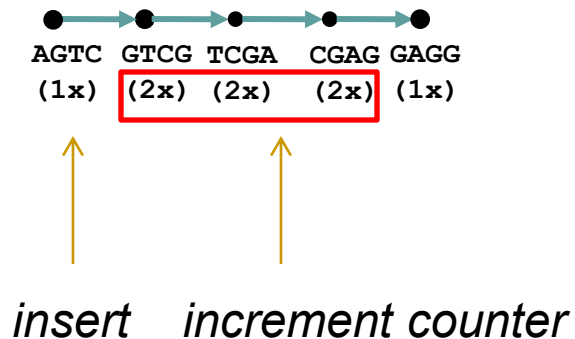




# A quick example

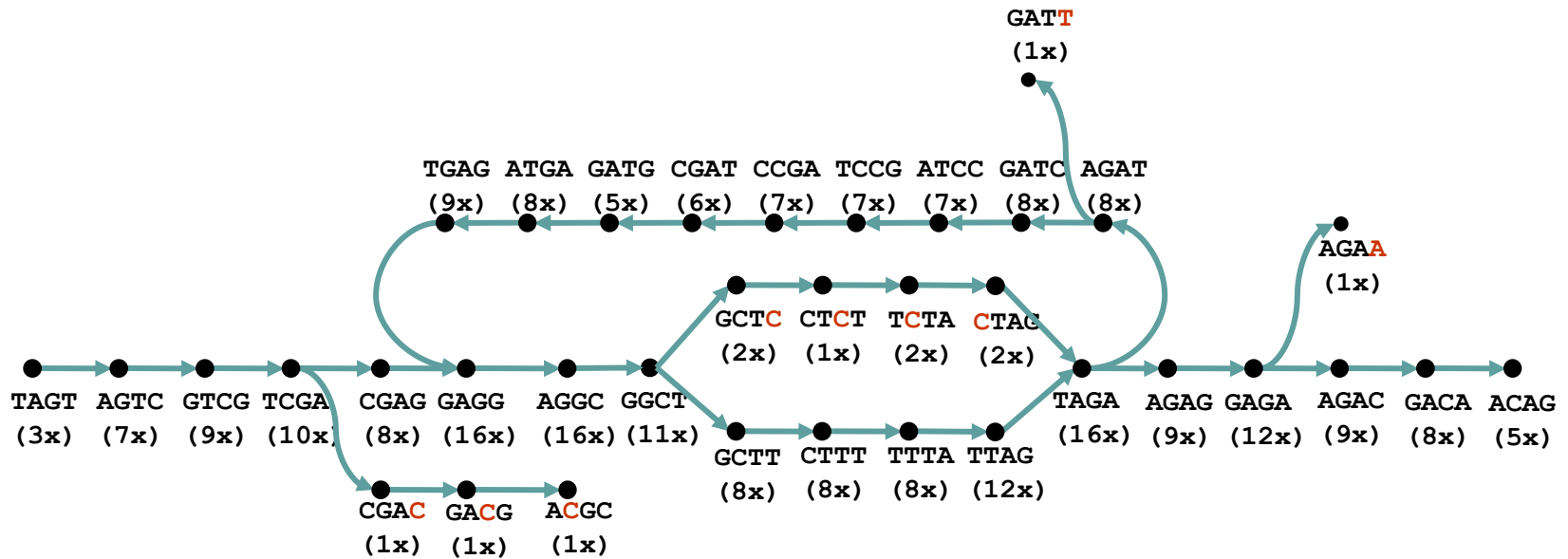
First read: **GTCGAGG**

Second read: **AGTCGAG**



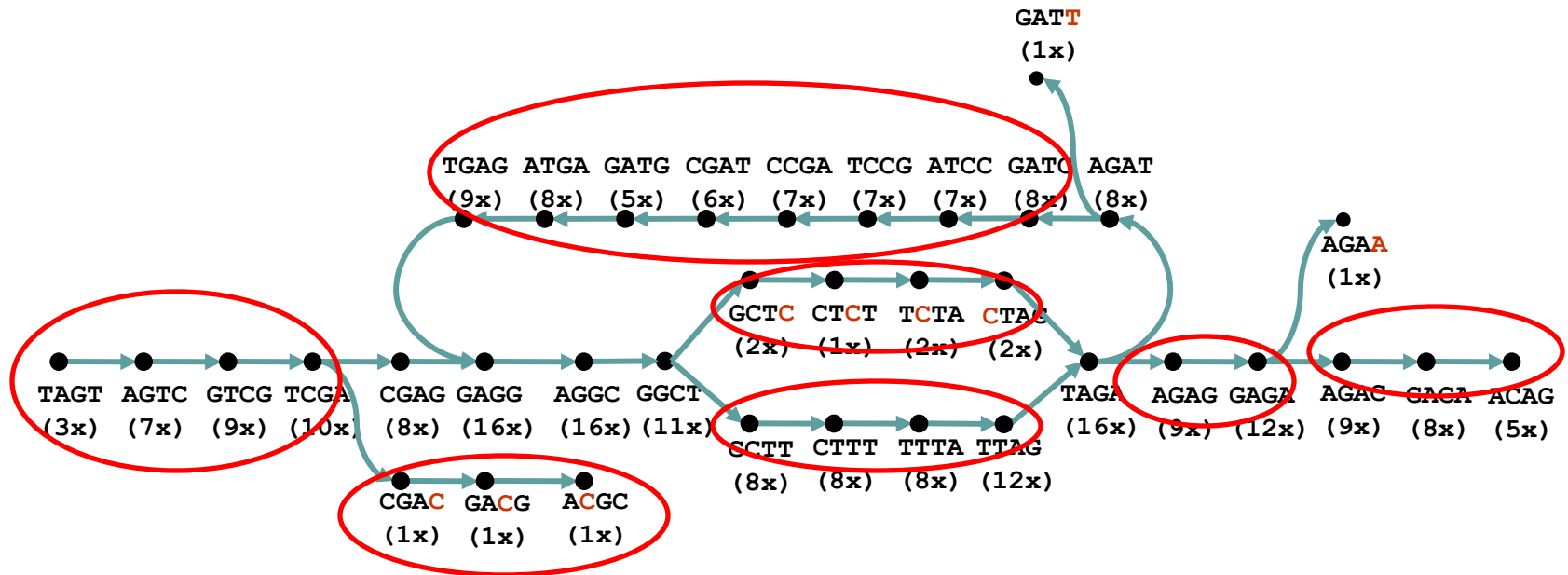
# A quick example

All the others...



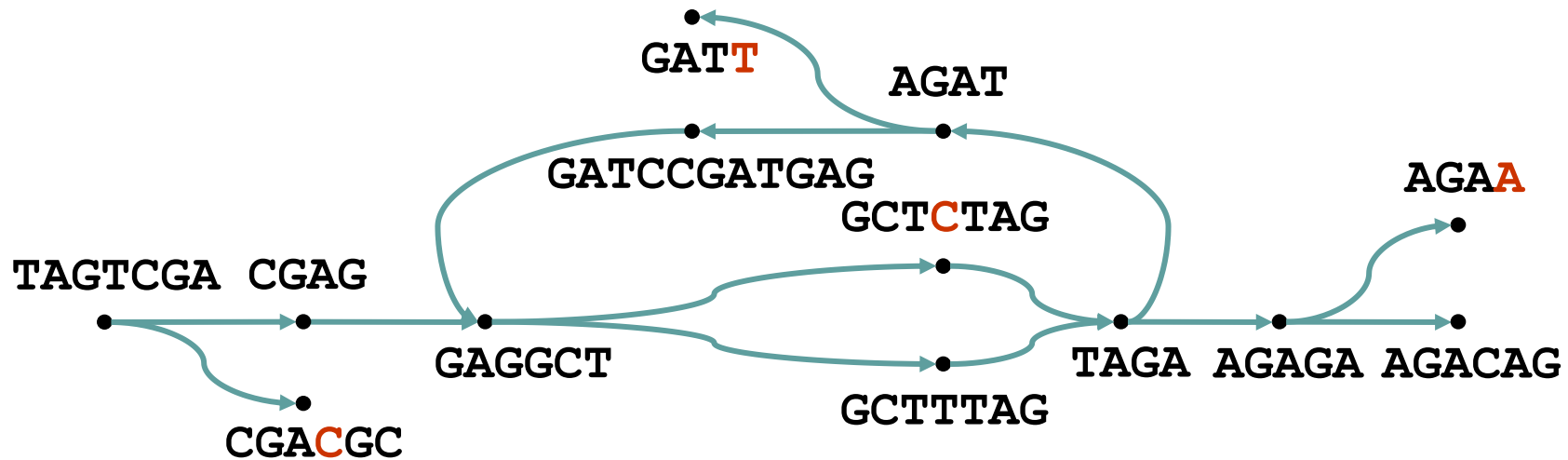
# A quick example

All the others...

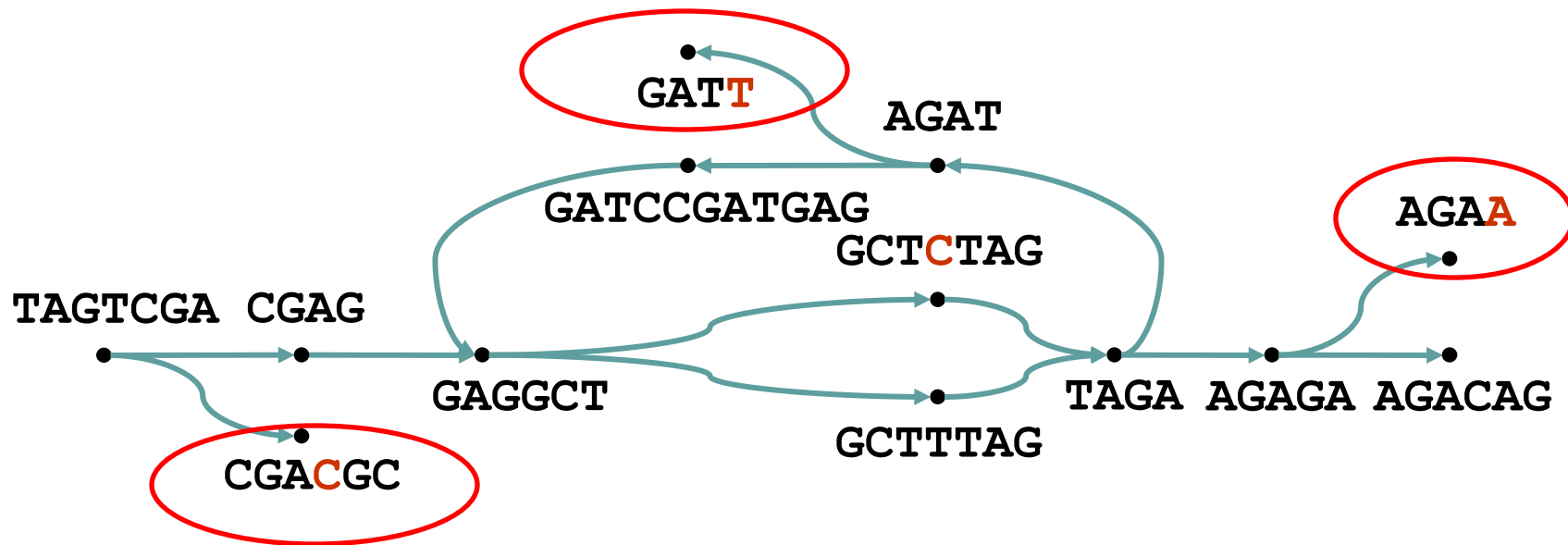


# A quick example

After simplification...

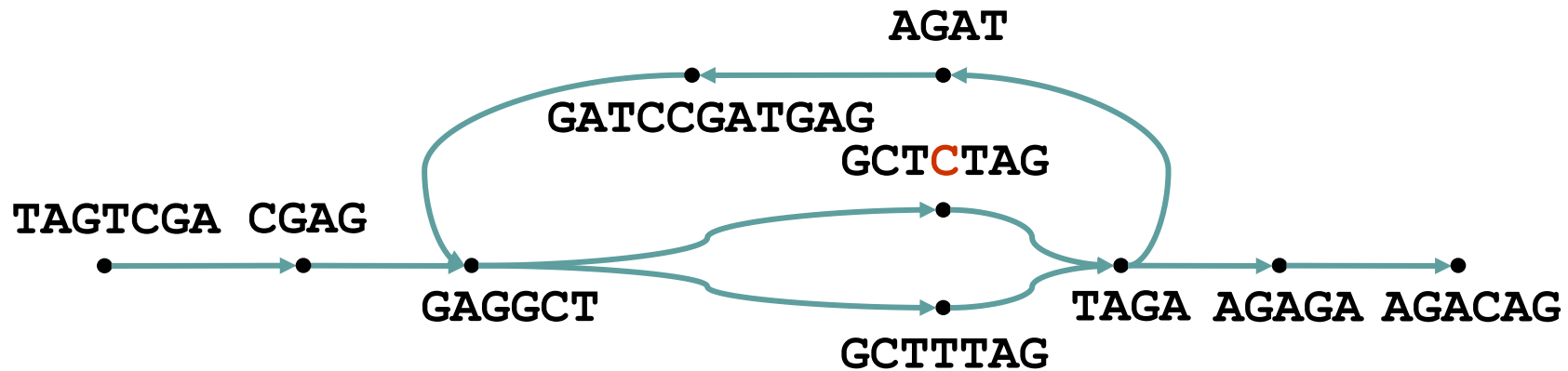


# Tips

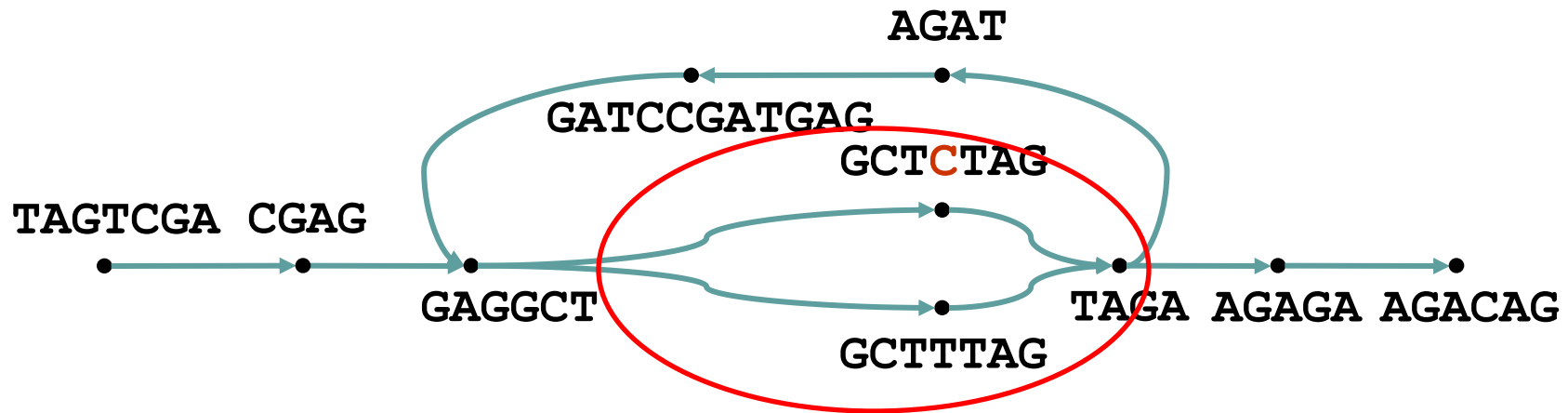


# Error removal

Tips removed...

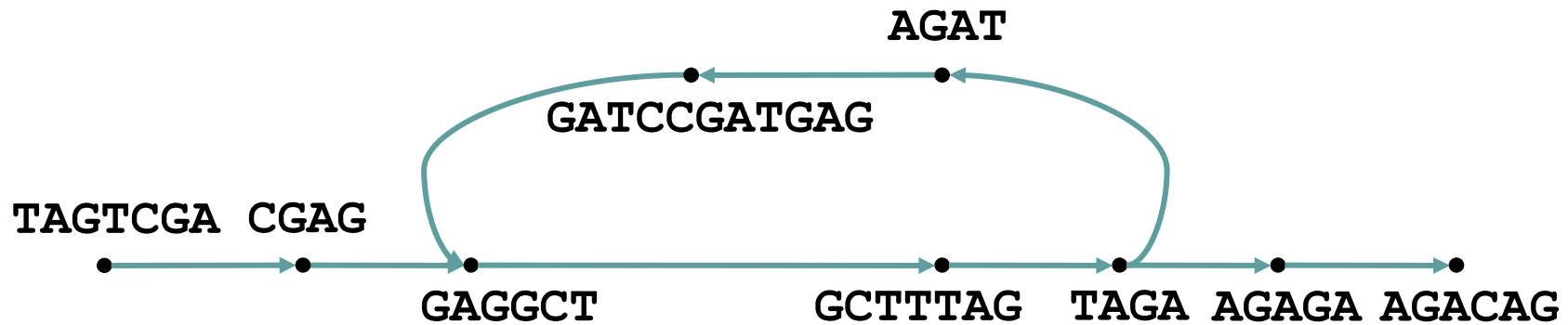


# Bubbles



# Error removal

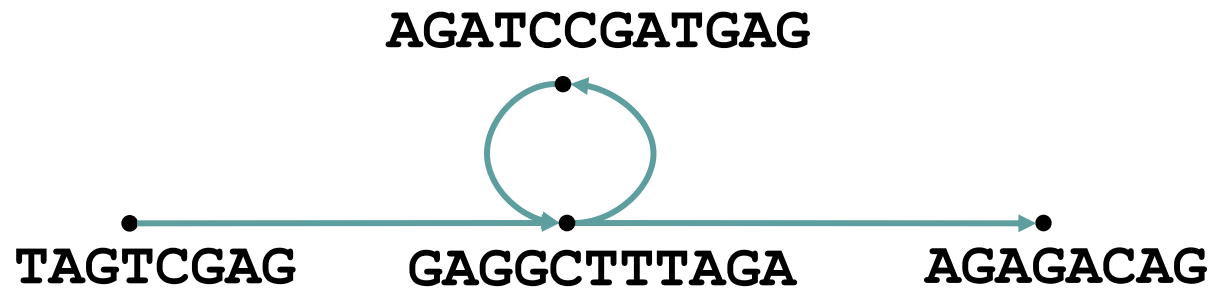
## Bubbles removed



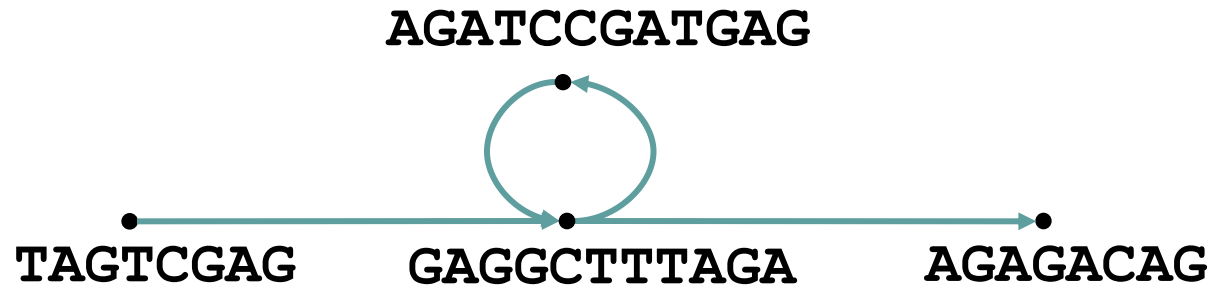


# Error removal

Final simplification...



# Eulerian path



TAGTCGAG **GAGGCTTTAGA** AGATCCGATGAG **GAGGCTTTAGA** AGAGACAG

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# Differences: de Bruijn vs Overlap

- Algebraic difference:
  - Reads in the OLC methods are atomic
  - Reads in the DB graph are sequential paths through the graph
- This leads to practical differences:
  - DB graphs allow for a greater variety of overlaps.
  - Overlaps in the OLC approach require a global alignment, not just a shared  $k$ -mer

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# Considerations

- Graph size scales with genome size
    - Increased error rate -> larger graph
  - Clipping to short k-mers get rid of sequence errors accumulated at the ends of reads
  - k value:
    - Small -> increased connectivity vs. more repeat collapses
    - Large -> increased specificity vs. decreased connectivity
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Resolving repeats using long reads or paired-end reads

# **REPEAT RESOLUTION**

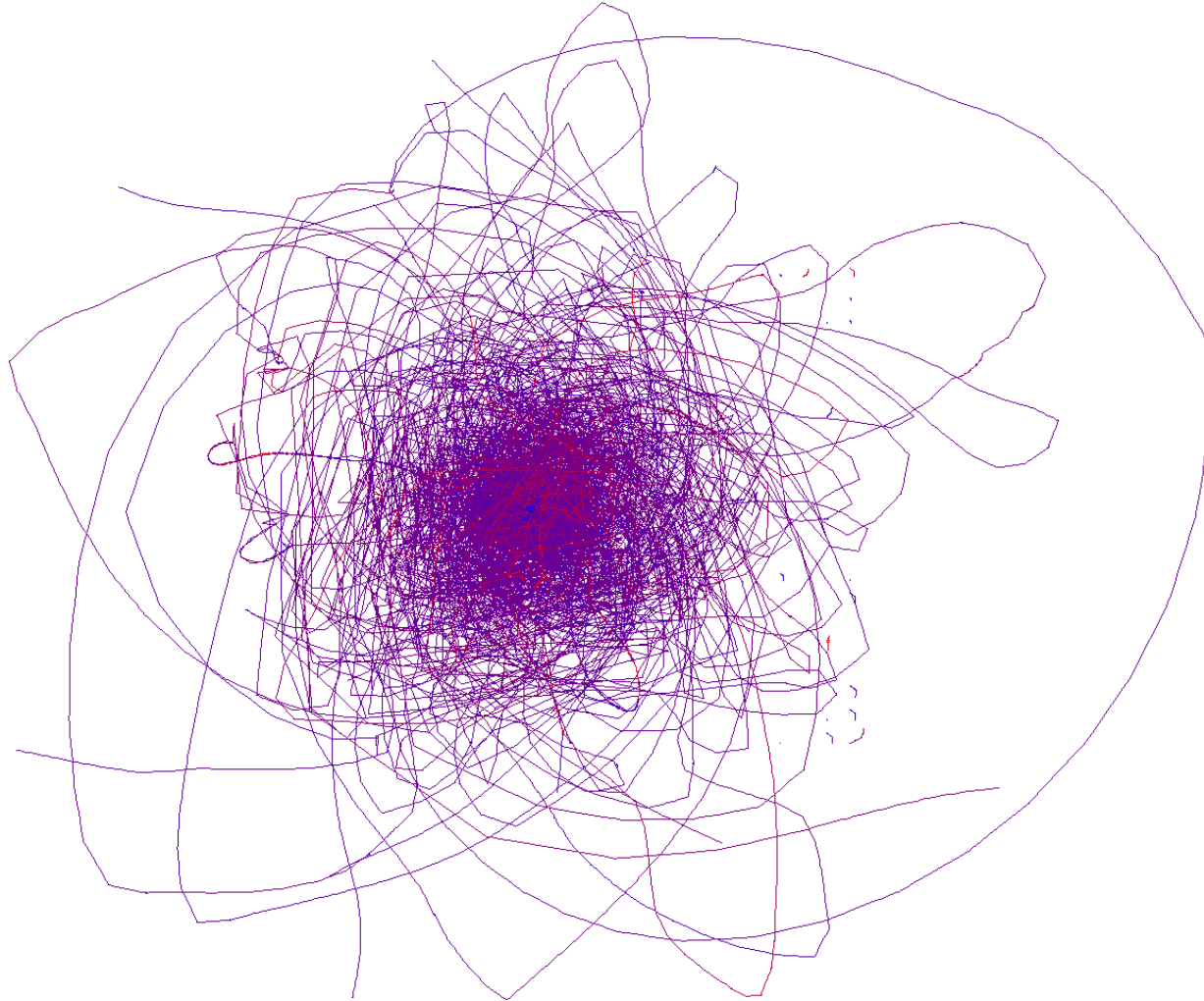
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# Chromosome X

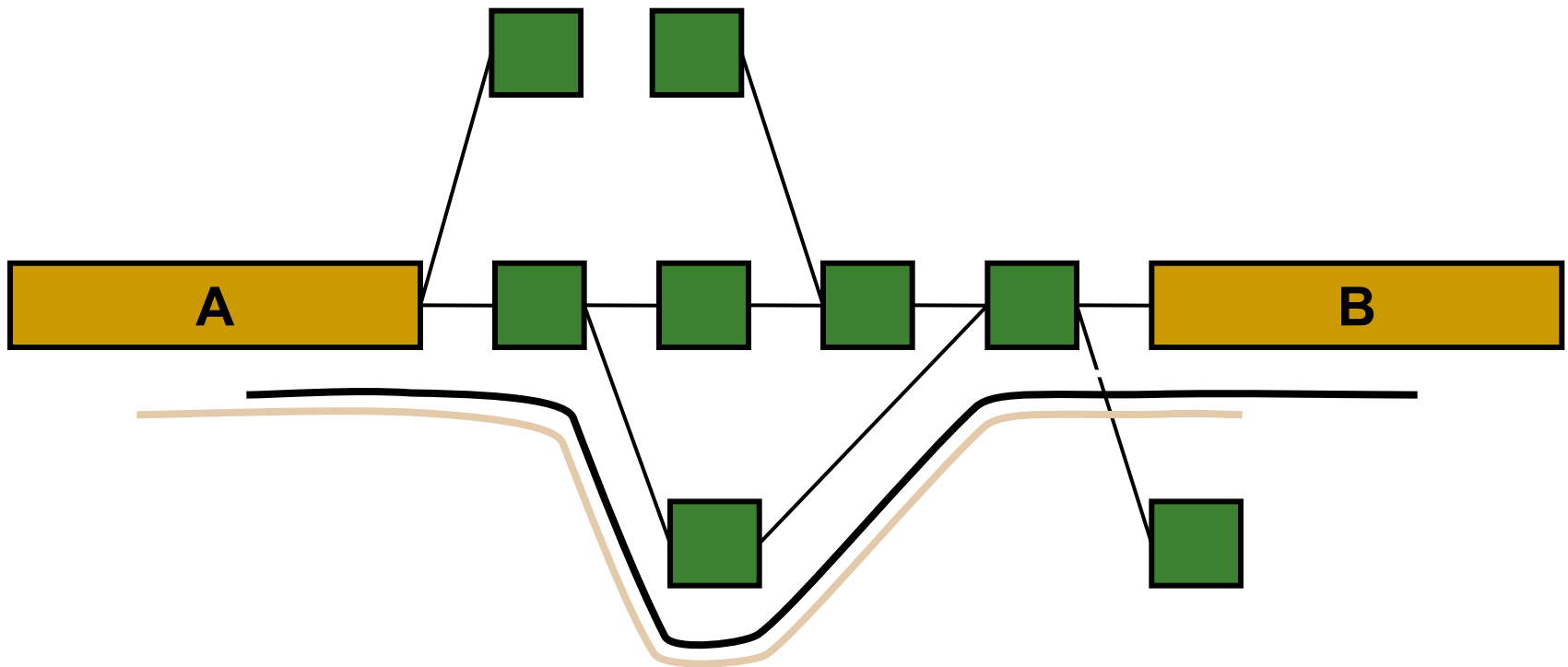
- 548 million Illumina reads were generated from a flow-sorted human X chromosome.
  - Fit in 70GB of RAM.
  - Many contigs: 898,401 contigs
  - Short contigs: 260bp N50 (max 6,956bp)
  - Overall length: 130Mb.
- Moral: there are engineering issues to be resolved but the complexity of the graph needs to be handled accordingly.
  - Reduced representation (Margulies et al.).
  - Combined re-mapping and de novo sequencing (Cheetham et al., Pleasance et al.).
  - Code parallelization (ABYSS)
  - Improved indexing (Cortex).
  - Use of intermediate re-mapping

# Repeats in a de Bruijn graph



*Slide courtesy of Dan Zerbino*

# Velvet: RockBand



Use long and short reads together



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# Different approaches to repeat resolution

- Theoretical: spectral graph analysis
  - Equivalent to a Principal Component Analysis
  - Relies on a (massive) matrix diagonalization
  - Comprehensive: all the data is integrated at once
  - Robust: small variations don't disturb the overall result
  - Never used because of the computational cost.

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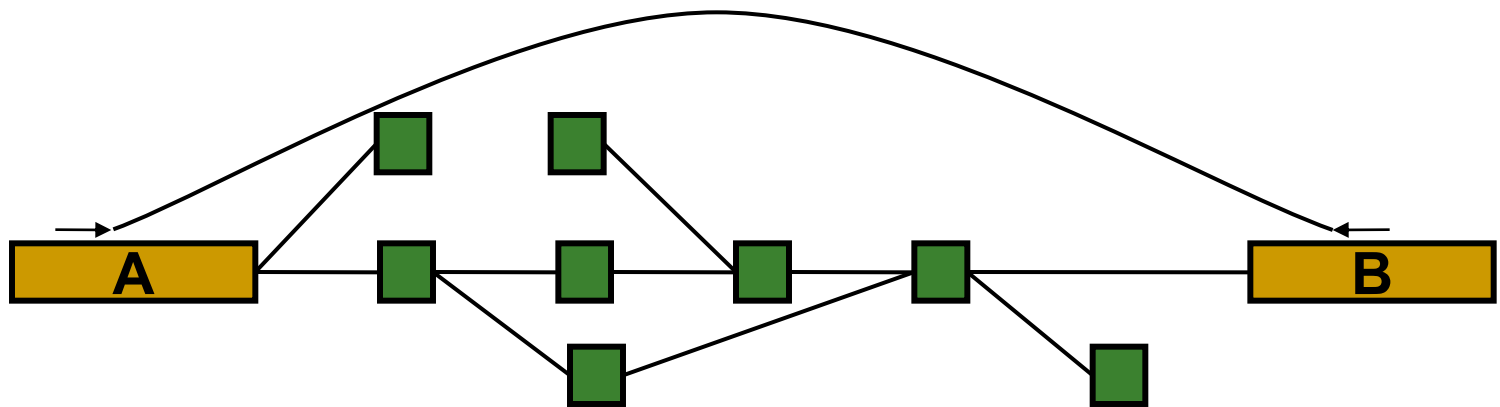
# Different approaches to repeat resolution

- Traditional scaffolding
  - e.g. Arachne, Celera, BAMBUS.
  - Heuristic approach similar to that used in traditional overlap-layout-consensus contigging.
  - Build a big graph of pairwise connections, simplify, extract obvious linear components.

# Different approaches to repeat resolution

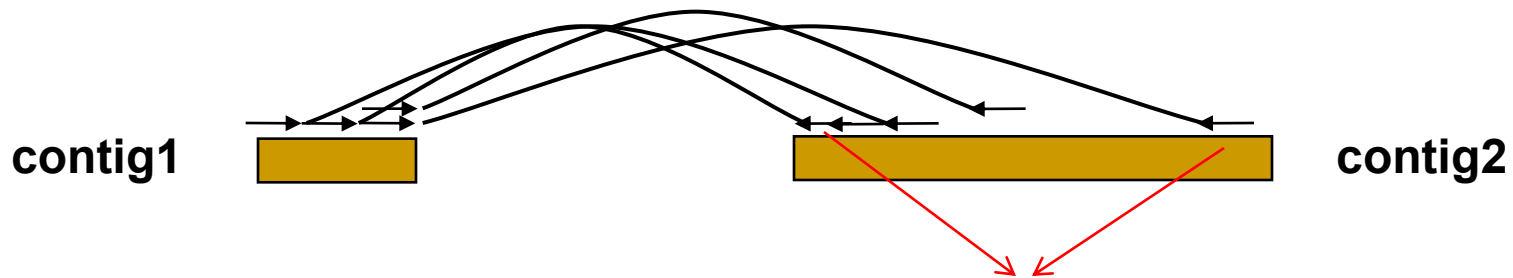
## ■ In NGS assemblers:

- EULER: for each pair of reads, find all possible paths from one read to the other.
- ABySS: Same as above, but the read-pairs are bundled into node-to-node connections to reduce calculations
- ALLPATHS: Same as above, but the search is limited to localized clouds around pre-computed scaffolds.



# Different approaches to repeat resolution

- Using the differences between insert length
  - The Shorty algorithm uses the variance between read pairs anchored on a common contig on  $k$ -mer.



*Collapsed repeat in contig1 ?*

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# **PRACTICAL CONSIDERATIONS**

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# Colorspace

- Di-base encoding has a 4 letter alphabet, but very different behavior to sequence space
  - Different rules for complementarity
- Direct conversion to sequence-space is simple but erroneous
  - One error messes up all the remaining basepairs
- Conversion must therefore be done at the very end of the process, when the reads are aligned
  - You can then use the transition rules to detect errors

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# Different error models

- When using different technologies, you have to take into account different technologies
  - Easy for OLC assembly
  - Much more tricky for de Bruijn assembly, since k-mers are not assigned to reads.
  - Different assemblers have different settings

# Pre-filtering the reads

- Some assemblers have built-in filtering of the reads (e.g. Euler) but not a generality.
  - Low phred quality
  - Reads with N characters
- Efficient filtering of low quality bases can cut down on the computational cost (memory & time)
- Some assemblers require reads of identical lengths.



